

LETTER



Alternative RNA splicing and alternative transcription start/end in acute respiratory distress syndrome

Alger M. Fredericks¹, Li Juan Wang², William G. Fairbrother³, Alfred Ayala¹ and Sean F. Monaghan^{1*} 

© 2020 Springer-Verlag GmbH Germany, part of Springer Nature

Dear Editor,

Critically ill patients develop acute respiratory distress syndrome (ARDS) [1] and despite the study of genomics of ARDS, there is little progress [2]. The drop in the cost of sequencing has refocused genetic studies from DNA to RNA sequencing and methods to analyze these data have improved [3]. The objective of this investigation is to utilize RNA sequencing data and analysis to identify novel gene targets in ARDS.

Methods are described in the eSupplement. The human cohort generated from the GTEx consortium consisted of 25 deceased patients with ARDS identified by the presence of diffuse alveolar damage (DAD, this limitation is described in the supplement), and 74 deceased patients evaluated to not have DAD. The mouse ARDS cohort included C57BL/6 mice ages 10–12 in a model previously described and compared to controls [4, 5].

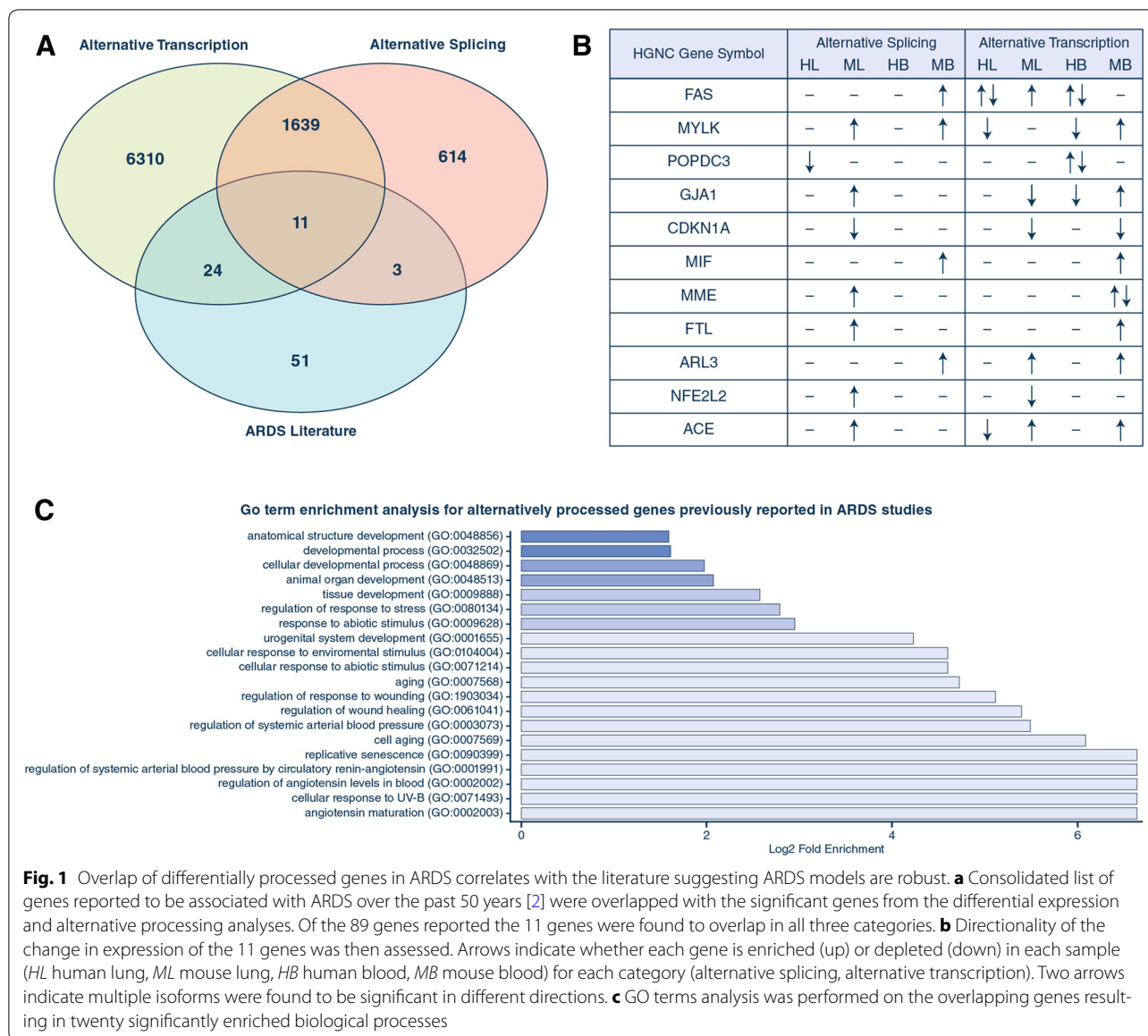
Alternatively spliced RNA arise from co/post-transcriptional events facilitated by the spliceosome, introns are removed to form the mature RNA from which protein isoforms are translated. Alternatively transcribed genes are the product of changes in promoter usage, polyadenylation signals, and RNA polymerase II interactions with DNA which can lead to changes in isoform usage similar to alternative splicing events. These are identified from the analysis of RNA sequencing data (see supplement). Significant differentially alternatively transcribed genes and alternative spliced genes were identified (see supplement) and were overlapped with genes previously

reported as ARDS related (Fig. 1a). Of 89 reported ARDS related genes [2], 38 were confirmed in at least one differential category confirming that our use of humans and mice with DAD/ARDS is appropriate and robust ($p = 1.25e-14$). Eleven previously reported genes were present in all categories (Fig. 1a). These 11 genes were evaluated for the change in alternative splicing and alternative transcription (Fig. 1b). GO term enrichment analysis was performed on the 11 overlapping genes, revealing 20 significant biological processes including ontology related to aging, and response to abiotic/environmental stimuli (Fig. 1c). There are 1639 genes that show overlap in alternative splicing and alternative transcription that were not previously in the literature (Fig. 1a). These genes were assessed for directionality alternative splicing and alternative transcription and GO terms (eTables 1, 2 in the supplement) should provide the foundation for future work in ARDS.

Studying the underlying changes in RNA processing (alternative splicing and alternative transcription start/end) not only expands basic knowledge of pathogenicity, but also provides additional targets for therapeutics. The most enriched GO term from the alternative splicing set (eFigure 1B), carboxy-terminal domain protein kinase complex (GO:0032806) refers to phosphorylation of the CTD of RNA polymerase II, which is vital in the regulation of transcription and RNA processing. In addition RNA polymerase complex binding (GO:0000993), and transport of the SLBP Independent/Dependent mature mRNA (R-HSA-159227; R-HSA-159230) are among the most enriched (eFigure 1B). This suggests alternative pre-mRNA splicing plays the dominant role in isoform usage in genes where expressions levels do not change, whereas alternative transcription may regulate isoform usage in

*Correspondence: smonaghan@lifespan.org

¹ Department of Surgery, Rhode Island Hospital, Brown University, 593 Eddy Street, Middle House 211, Providence, RI 02903, USA
Full author information is available at the end of the article



genes that are more dynamically expressed during critical illness. Although it is possible the enrichment reflects down regulation through inhibitory genes, these data support the hypothesis that alternative splicing and alternative transcription may have separate roles in DAD/ARDS by regulating different genes to perform distinctive functions.

In this analysis of RNA sequencing data from deceased patients with ARDS identified by the presence of DAD and a clinically relevant mouse model of ARDS, novel genes were identified. Future research is needed using on the mechanism of alternative RNA splicing and alternative transcription start/end seen in ARDS.

Electronic supplementary material

The online version of this article (<https://doi.org/10.1007/s00134-020-05953-3>) contains supplementary material, which is available to authorized users.

Author details

¹ Department of Surgery, Rhode Island Hospital, Brown University, 593 Eddy Street, Middle House 211, Providence, RI 02903, USA. ² Department of Pathology, Rhode Island Hospital, Brown University, 593 Eddy Street, Providence, RI 02903, USA. ³ MCB Department, Brown University, 70 Ship Street, Room 4040, Providence, RI 02903, USA.

Acknowledgements

This work was funded in part by NIH-NIGMS P20GM103652 (S. F. M.); NIH-NIGMS R35GM118097 (A. A.), the C. James Carrico, MD, FACS, Faculty Research Fellowship for the Study of Trauma and Critical Care from the American College of Surgeons (S. F. M.); NIH-NIGMS R01GM127472 (W. G. F.)

Author contributions

AMF and SFM had full access to all of the data in the study and took responsibility for the integrity of the data and the accuracy of the data analysis. Concept and design: AMF, WGF, AA, SFM. Acquisition, analysis, or interpretation of data: AMF, LJW, WGF, AA, SFM. Drafting of the manuscript: AMF, AA, SFM. Critical revision of the manuscript for important intellectual content: AMF, LJW, WGF, AA, SFM. Statistical analysis: AMF, SFM. Obtained funding: WGF, AA, SFM. Administrative, technical, or material support: AMF, LJW. Supervision: WGF, AA, SFM.

Compliance with ethical standards**Conflicts of interest**

The authors declare that they have no competing interests.

Publisher's Note

Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.

Accepted: 26 January 2020

Published online: 20 February 2020

References

1. Thompson BT, Chambers RC, Liu KD (2017) Acute respiratory distress syndrome. *N Engl J Med* 377:562–572
2. Reilly JP, Christie JD, Meyer NJ (2017) Fifty years of research in ARDS. Genomic contributions and opportunities. *Am J Respir Crit Care Med* 196(9):1113–1121
3. Stark R, Grzelak M, Hadfield J (2019) RNA sequencing: the teenage years. *Nat Rev Genet* 20(11):631–656. <https://doi.org/10.1038/s41576-019-0150-2>
4. Ayala A, Chung CS, Lomas JL, Song GY, Doughty LA, Gregory SH, Cioffi WG, LeBlanc BW, Reichner J, Simms HH, Grutkoski PS (2002) Shock-induced neutrophil mediated priming for acute lung injury in mice: divergent effects of TLR-4 and TLR-4/FasL deficiency. *Am J Pathol* 161:2283–2294
5. Lomas-Neira JL, Heffernan DS, Ayala A, Monaghan SF (2016) Blockade of endothelial growth factor, angiopoietin-2, reduces indices of ARDS and mortality in mice resulting from the dual-insults of hemorrhagic shock and sepsis. *Shock (Augusta, GA)* 45:157–165